

# EVALUATION ON THE EFFICACY OF SOME SELECTED BOTANICALS IN CONTROLLING THE COTTON MEALYBUG *PHENACOCCLUS SOLENOPSIS* (TINSLEY)

M. Prishanthini and M. Vinobaba

Department of Zoology, Faculty of Science,  
Eastern University, Sri Lanka

## Introduction

The mealybug species are widespread throughout the world. The cotton mealybug *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae) has been described as a serious and invasive polyphagous pest with a vast host range by several authors. It has a wide geographical distribution with its origin in Central America (Fuchs et al., 1991) followed by reports of the Caribbean and Ecuador (Ben-Dov, 1994), Chile (Larrain, 2002), Argentina (Granara de Willink, 2003), Brazil (Mark and Gullan 2005). *P. solenopsis* has been described as a serious and invasive pest of shoe flower in Pakistan and India (Hodgson et al. 2008) on *Hibiscus rosa-sinensis* in Nigeria (Akintola and Ande, 2008). Latest report by the authors on the invasiveness of *P. solenopsis* has been from the Eastern region of Sri Lanka (Prishanthini and Vinobaba, 2009) on ornamentals, vegetable crops, and weeds, and in China (Wang et al. 2009; Wu and Zhang, 2009) on shoe flower. Reliance on synthetic chemicals to control pests has also given rise to a number of problems such as destruction of beneficial non-target organisms (parasitoids and predators) and can lead to secondary outbreaks of pests that are normally under natural control resulting in their rapid proliferation. Scientists are now experimenting and working to protect insect infestation by indigenous plant materials (Roy et al., 2005). There are many research evidences on the application of botanical pesticides against various mealybugs. Therefore present study conducted to evaluate the efficacy of some of the selected, locally available botanical pesticides in controlling mealybugs in the homegardens.

## Methodology

The botanicals used in this experiment were, *Ocimum sanctum* L. (Lamiaceae), *Azadirachta indica* A. Juss. (Meliaceae), *Calotropis gigantea* R. Br. (Asclepidaceae), *Nicotina tabacum* Linn. (Solanaceae) and *Alium sativum* Linn. (Amaryllidaceae). Leaves of *Ocimum sanctum* (Tulasi) and *Azadirachta indica* (Neem) and *Calotropis sp.* were collected from the homegardens of Batticaloa. The dried leaves of Tobacco (*Nicotina tabacum*) and Garlic cloves were bought from local market. The leaves collected were cleaned and allowed to dry under solar radiation for one week. Then the dried materials were powdered and stored separately in dark bottles for extraction. Extracts of the 50 g of powdered botanicals were prepared using ethanol extraction method. Dried extracts were weighed and the stock solutions were prepared. Further dilutions were made with 0.2, 0.4, 0.6, 0.8, 1.0, 1.2 and 1.5 percentages concentrations. Dilutions prepared were applied over approximately 10-12 cm length terminal portions of the Shoe flower stems (*Hibiscus rosa-sinensis*) infected with the *P. solenopsis* adults. The mortality records were obtained in percentage values. The most effective botanical solution which has the lowest LC<sub>50</sub> (concentration which cause lethality of 50% of the mealybug population) was used for the field trial. Shoe flower plants (20 Nos) of 50- 60 cm height were infected with 20 adult female mealybugs and after 35 days of development the numbers of adult females, and nymphs were counted using hand lens. Solution was prepared and applied over the mealybug colonies and the mealy bug mortality was counted at 24, 48 and 72 hours after initial application. For all the data obtained the

differences among the mortality of mealybugs at all treatments were subjected to analysis of variance (one way ANOVA) and differences among means were considered significant at a probability level of five percent ( $p \leq 0.05$ ). Probit analysis was carried out to determine the  $LC_{50}$  values of each botanical. Statistical package Minitab 14.0 was used for all these statistical analyses.

### Discussion and Conclusion

The results revealed that the treatments are significantly differing among themselves in causing mortality of *P.solenopsis*. ( $p < 0.05$ ) except at 0.2% concentration ( $p = 0.230$ ). Figure 1 shows the comparison of percentage mean mortalities to different botanicals at different concentrations at 24 hrs after initial applications. Mortality rates increased with increasing concentrations for all botanicals  $LC_{50}$  values obtained from probit analysis for mortality values after 24 hours of each botanical applied are given in the table 1. According to the results of probit analysis the botanical *O.sanctum* has the lowest value i.e. 0.60 % solution. Among all, the *Ocimum sanctum* and soap mixed solution was found to be more effective against *P.solenopsis* with the lowest  $LC_{50}$  value.

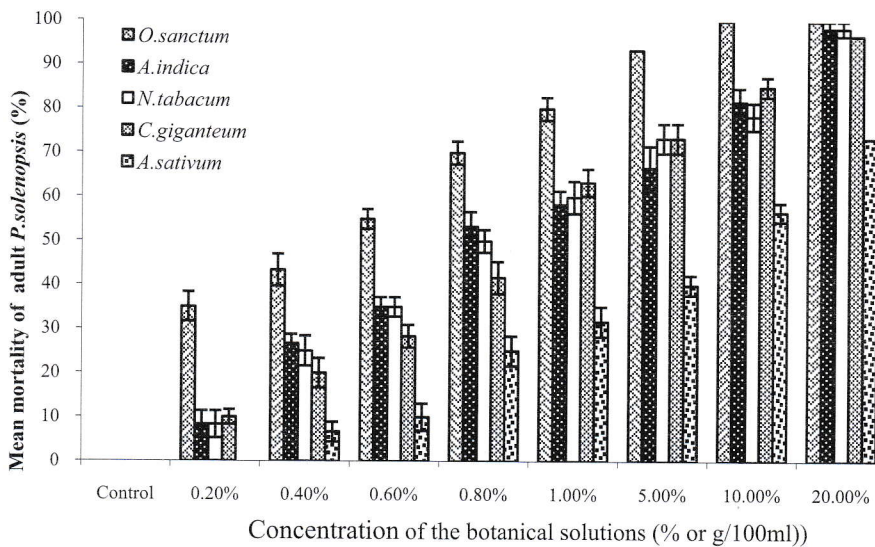
In the field experiment the 0.6% solution of *Ocimum sanctum* which is the  $LC_{50}$  value obtained from the laboratory experiments was field applied on healthy shoe flower plants of same age and height. In this trial the 0.6% Tulasi, *Ocimum sanctum* solution caused 39.42% adult mortality and 72.21% nymphal mortality to *P.solenopsis*. Adult mortality under field conditions was lower than that of under laboratory conditions. Use of Tulasi for pest control has long history and has both repellent and herbicidal properties. The essential oils from the species of this genus contain linalool, linalol, linoleic acid, *p*-cymene, estragol, eucalyptol, eugenol, citral, thujone, ocimene, camphor, methyl chavicol, oleic acid, and many other terpenes as active ingredients, all of which are effective repellents (Moore and Lenglet, 2004). Generally contact pesticides are less effective against mealybugs because of their cryptic habitats in plants and the water proof waxy layer over the body (Tanwar *et al*, 2007). The soap solution added at low concentration increases the effectiveness of the botanical solution. Soap facilitates the solubility of the active ingredient and acts as a sticking agent (Nhachi and Kasilo, 1996), breaks down the protective wax cover and also acts as a surfactant.

Table 1:  $LC_{50}$  values (after 24 hrs) for the botanicals obtained from Probit Analysis

Name of Botanical	$LC_{50}$
<i>O.sanctum</i>	0.60
<i>N.tabacum</i>	0.89
<i>C.gigantea</i>	0.95
<i>A.sativum</i>	1.15
<i>A.indica</i>	0.82

As a conclusion it can be stated that *O.sanctum* was effective significantly at lower concentrations and 0.6% concentration of the *O.sanctum* solution was resulted a significant nymphal mortality in the field conditions. The botanicals used in this study such as *A.indica*, *O.sanctum*, *C.gigantea*, *N.tabacum* and *A.sativum* were showed different levels of insecticidal activities.





Based on these results development of new formulations with the combinations of these botanicals which can be produced and applied using simple methods applicable to local public will be very useful. Moreover, analyzing new botanicals from different plant origins and least toxic chemicals for their efficacy against the mealybugs are also necessary to reduce use of the toxic chemical insecticides. More research on the active ingredients, pesticide preparations, application rates and environmental impact of botanical pesticides are a prerequisite for sustainable agriculture.

## References

- Akintola AJ and Aude AT. 2008. First Record of Phenacoccus solenopsis Tinsley (Hemiptera: Pseudococcidae) on Hibiscus rosa-sinensis in Nigeria. Agricultural Journal (Medwell Journals, Pakistan) 3(1): 1-3.
- Ben-Dov Y. 1994. A systematic catalogue of the mealybugs of the world, p.686. Intercept Limited, Andover, UK.
- Fuchs TW, Stewart JW, Minzenmayer R and Rose M. 1991. First record of Phenacoccus solenopsis Tinsley in cultivated cotton in the United States. Southwestern Entomologist 16(3): 215-221.
- Williams DJ and Granara de Willink MC. 1992. Mealybugs of Central and South America, p. 635. CAB International.
- Hodgson CJ, Abbas G, Arif MJ, Saeed S and Karar H. 2008. Phenacoccus solenopsis Tinsley (Sternorrhyncha: Coccoidea: Pseudococcidae), an invasive mealybug damaging cotton in Pakistan and India, with a discussion on seasonal morphological variation. Zootaxa 1913: 1-35.
- Larrain SP. 2002. Insect and mite pest incidence on sweet pepinos Solanum muricatum (Ait.) cultivated in the IV Region, Chile. Agricultura-Tecnica 62(1): 15-26.
- Moore, S. J. and Lenglet, A. D. (2004). An Overview of Plants Used as Insect Repellents, Traditional Medicinal Plants and Malaria pp-343-359. Available from-[http://sjmoore.net/doc/Overview\\_of\\_plant\\_based\\_repellents.pdf](http://sjmoore.net/doc/Overview_of_plant_based_repellents.pdf) Accessed on 5/7/2009

- Nhachi, C. F. B and Kasilo O. M. J. (1996). Pesticides in Zimbabwe: Toxicity and health implications. University of Zimbabwe publications, Harare
- Prishanthini, M. and Vinobaba, M. (2009), First record of new exotic Mealybug species, *Phenacoccus solenopsis* Tinsley (Hemiptera: Pseudococcidae), its Host range and abundance in the Eastern Sri Lanka, *Journal of Science* 6(1),88-100
- Roy B., Amin R., Uddin M. N., Islam A. T. M. S., Islam M. J. and Halder, B. C. (2005). Leaf extracts of *Shiyalmutra* (*Blumea lacera* Dc.) as botanical pesticides against lesser grain borer and rice weevil. *Journal of Biological Sciences* 5 (2), 201 – 204.
- Tanwar, R. K, Jeyakumar, P. and Monga, D. (2007). Mealybugs and their management. Technical Bulletin 19, September, 2007, National Center for Integrated Pest Management, Pusa Campus, New Delhi. India Available from: <http://www.ncipm.org.in> (Accessed on: 18 March 2009)
- Wang YP, Wu SA and Zhang RZ. 2009. Pest risk analysis of a new invasive pest *Phenacoccus solenopsis*, to China. (in Chinese; Summary in English). *Chinese Bulletin of Entomology* 46(1):101-106.
- Weinberger, R. A. (2005), Botanical insecticides, soaps and oils, In: *Biological and Biotechnological control of pests*, CAB international, Wallingford. Oxon.pp-114-115.
- Wu SA and Zhang RZ. 2009. A new invasive pest, *Phenacoccus solenopsis* threatening seriously to cotton production. (in Chinese; Summary in English). *Chinese Bulletin of Entomology* 46(1): 159-162.